

## IN THE UNITED STATES PATENT AND TRADEMARK OFFICE

Attorney Docket No.: P-576 (TI-0022)  
Inventors: Huber et al.  
Serial No.: 09/770,410  
Filing Date: January 25, 2001  
Examiner: Therkorn, Ernest G.  
Customer No.: 26259  
Group Art Unit: 1723  
Confirmation No.: 6186  
Title: Method and Apparatus for Separating  
Polynucleotides Using Monolithic  
Capillary Columns

Electronically Submitted via EFS-Web

Date: September 5, 2006

I hereby certify that this paper is being electronically  
submitted on the date indicated above to the  
Commissioner for Patents, U.S. Patent &  
Trademark Office.

By Jane Massey Licata  
Typed Name: Jane Massey Licata, Reg. No. 32,257

Commissioner for Patents  
U.S. Patent & Trademark Office

Dear Sir:

## DECLARATION UNDER RULE § 1.131

We, Christian Huber, Herbert Oberacher and Andreas  
Premstaller, hereby declare that:

1. We are co-inventors in U.S. Patent Application  
Serial No. 09/770,410 filed June 7, 2000 and are most familiar  
with the subject matter of this application and the research  
effort which lead to the discovery of the instant invention. All  
the work described in the following paragraph occurred at the  
Institute of Analytical Chemistry and Radiochemistry in

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Innsbruck, Austria, a recognized WTO member country since January 1, 1995.

2. We have reviewed Gusev et al. ((September 1999) *J. Chromatography* 855:273-290) and find that this reference describes a porous monolithic packing prepared with polystyrene-divinylbenzene support which is covalently attached to a fused silica capillary inner wall treated with a coupling agent trimethoxysilyl propyl methacrylate to provide anchoring sites for grafting of the polymer to the silica surface. The median pore radius for a monolithic sample prepared with ethanol is, as estimated by Gusev, about 5 micrometers.

3. Our invention referenced above, teaches a device for separating a mixture of polynucleotides by ion pair-reversed phase-high performance liquid chromatography. The device comprises a polymeric monolith having non-polar chromatographic surfaces. The monolith comprises an underivatized poly-(styrene/divinylbenzene) matrix and is contained within a tube having an inner diameter in the range of 1 to 1000 micrometers.

4. Laboratory protocol notebooks regarding experiments related to this invention were kept by Andreas Premstaller as a Ph.D. student under the direction of Christian Huber.

5. Andreas Premstaller worked in Christian Huber's laboratory during 1998 and 1999.

6. According to laboratory protocol notebooks submitted herewith, the first synthesis of PS/DVB monolith using decanol and tetrahydrofuran as porogens was performed on August 6, 1998. We then succeeded in a first separation of proteins (lysosome from beta-lactoglobulin B) in a PS/DVB monolithic column on August 25, 1998. See, e.g., the chromatograph at the bottom right-hand corner of the fourth laboratory notebook page.

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The first successful separation of oligonucleotides on a PS/DVB monolith synthesized with decanol/THF as porogens was February 9, 1999.

7. We were able to fully practice our invention described in the above-referenced patent application prior to the date of the publication of the Gusev paper. A copy of the relevant laboratory notebook pages hereby accompanies my declaration.

We further declare that all statements made herein of our own knowledge are true and that all statements made on information and belief are believed to be true, and further, that these statements were made with the knowledge that willful false statements and the like so made are punishable by fine or imprisonment, or both, under Section 1001 of Title 18 of the United States Code, and that such willful false statements may jeopardize the validity of the application or any patent issuing thereon.

August 30, 2006  
Date

Christian Huber  
Christian Huber, Ph.D.

\_\_\_\_\_  
Date

\_\_\_\_\_  
Herbert Oberacher, Ph.D.

\_\_\_\_\_  
Date

\_\_\_\_\_  
Andreas Premstaller, Ph.D.

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\_\_\_\_\_  
Date

\_\_\_\_\_  
Christian Huber, Ph.D.

30.8.2006

\_\_\_\_\_  
Date

Herbert Oberacher  
\_\_\_\_\_  
Herbert Oberacher, Ph.D.

\_\_\_\_\_  
Date

\_\_\_\_\_  
Andreas Premstaller, Ph.D.

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The first successful separation of oligonucleotides on a PS/DVB monolith synthesized with decanol/TF as porogens was February 10, 1999.

7. We were able to fully practice our invention described in the above-referenced patent application prior to the date of the publication of the Gusev paper. A copy of the relevant laboratory notebook pages hereby accompanies my declaration.

We further declare that all statements made herein of our own knowledge are true and that all statements made on information and belief are believed to be true, and further, that these statements were made with the knowledge that willful false statements and the like so made are punishable by fine or imprisonment, or both, under Section 1001 of Title 18 of the United States Code, and that such willful false statements may jeopardize the validity of the application or any patent issuing thereon.

\_\_\_\_\_  
Date

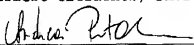
\_\_\_\_\_  
Christian Huber, Ph.D.

\_\_\_\_\_  
Date

\_\_\_\_\_  
Herbert Oberacher, Ph.D.

25.07.2006

\_\_\_\_\_  
Date

  
\_\_\_\_\_  
Andreas Premstaller, Ph.D.

58.18

Umschmelzung mit THF

CaO<sub>2</sub>  
mischelöslich  
7.88

Nr.	Datum	Kapillare	Polymerisationsmischung					Temperatur [°C]
			Styrol [ml]	DVB [ml]	AIBN [g]	C12OH [ml]	THF [ml]	
M14_1	05.08.98	320/450 WBH06A, 20 cm, vs	1,00	1,00	0,050	3,00	0,00	70, TS
M14_2	05.08.98	320/450 WBH06A, 20 cm, vs	1,00	1,00	0,050	2,80	0,10	70, TS
M14_3	05.08.98	320/450 WBH06A, 20 cm, vs	1,00	1,00	0,050	2,80	0,20	70, TS
M14_4	05.08.98	320/450 WBH06A, 20 cm, vs	1,00	1,00	0,050	2,70	0,30	70, TS
M14_5	05.08.98	320/450 WBH06A, 20 cm, vs	1,00	1,00	0,050	2,60	0,40	70, TS

THF sollte heraus in das Messen ein bisschen als Toluol  
THF distilliert, da mit Rohöl/Feig (Theil) stabilisiert

Ausgangsmaterial: VS 3.8.98 trocken  
THF dest.

Zeit: 6.8.98

T = 70°C

Zeit: 7.8.98

T = 70°C

7.8.98

M14_1 16 cm			M14_2 16 cm		
Fluss [l/min]	Gegensdruck [bar]	Reaktor [l/min]	Fluss [l/min]	Gegensdruck [bar]	Reaktor [l/min]
5	1	0,08	10	1	0,07
10	1	0,08	20	4	0,27
15	4	0,25	30	8	0,45
20	7	0,44	100	14	0,88
100	14	0,45	150	20	1,35
125	21	1,21	200	25	1,67
200	28	1,75			
k-Bar cm <sup>-1</sup> [l/min]			k-Bar cm <sup>-1</sup> [l/min]		
0,003712			0,003313		

M14_3 16 cm			M14_4 16 cm		
Fluss [l/min]	Gegensdruck [bar]	Reaktor [l/min]	Fluss [l/min]	Gegensdruck [bar]	Reaktor [l/min]
5	1	0,07	5	9	0,28
10	3	0,20	10	14	0,88
15	6	0,40	20	22	2,00
100	11	0,73	60	08	4,25
100	14	0,35	100	120	7,80
200	19	1,27	150	180	11,25
k-Bar cm <sup>-1</sup> [l/min]			k-Bar cm <sup>-1</sup> [l/min]		
0,004914			0,074816		

M14_5 16 cm			300 bar Gegenstand	
Fluss [l/min]	Gegensdruck [bar]	Reaktor [l/min]	Fluss [l/min]	Wegpunkt [l/min]
2	21	1,31	0,0%	0,00111
3	33	2,06	3,2%	0,00200
4	49	3,05	6,7%	0,00311
5	65	3,94	10,0%	0,07140
7	77	4,61	12,2%	0,08015
10	92	5,25		
15	170	10,53		
20	194	11,89		
k-Bar cm <sup>-1</sup> [l/min]			0,06148	

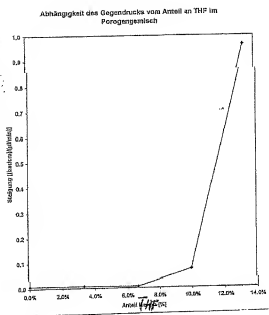
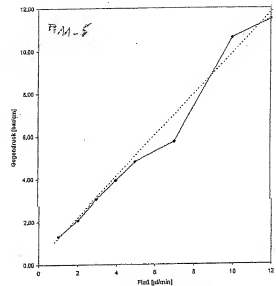
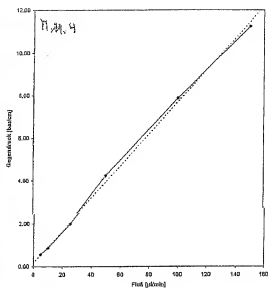
Umschmelzung: 55 Stk. 450 cm 1.8.98 → 8.98  
M14\_1 wurde 7.8.98  
sch. porös.

M14\_2 große Por., nicht gleichmäßig  
M14\_3 große Por., wurde 7.8.98 (sch. porös)

M14\_4 sch. dicht, keine Por., por., da nicht  
ganz zu Ende

M14\_5 keine Umstrukturierung zu erkennen.

ab



Als nächstes Band wurde DM.4 und DM.5 gegeben. Außerdem.

für alle DM.4 und DM.5 taken.

25.08.98

M 11.5 min 6.8.98

100%  $H_2O$  240 bar / 5  $\mu L/min$

File: AP80875.S170  
GKN-50FF

SYNAP, 130  $\mu L/min$   $\rightarrow$  Split  $\rightarrow$  4.6  $\mu L/min$   
2 min 15 sec / 10  $\mu L$   
2 min 30 sec / 4  $\mu L/min$

Gradient:

(A)  $H_2O$  0.1% TFA

(B) ACN 0.1% TFA

50% A, 14.50 -

Eluent - T-Held statt negative T-Held

10  $\mu L$ , 2 min 15 sec  $\frac{10}{2.75}$  4.44  $\mu L/min$

Reinigung:

Thrombin 0.05% in  $H_2O$  50% ACN, 0.1% TFA

pH 2.00 bar

100%  $H_2O$ , 0.1% TFA: Protein zeigt kein Peak  $\rightarrow$  keine Aktivität?

Thrombin near co. 1.50 min

50% ACN, 0.1% TFA: Protein fließt mit Thrombin: keine Retention  
RTB A.

27.8.98

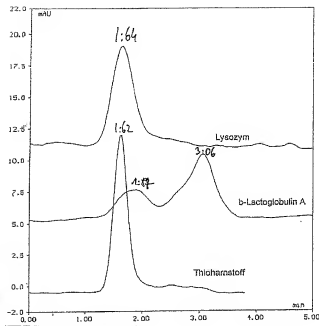
50% ACN, 0.1% TFA: Protein quickly eluted als Peak.

LAC A.

Lys kein Retention

Nun 40% ACN



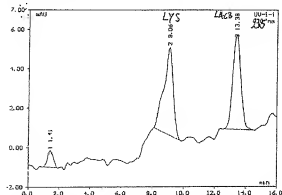


50% ACN, 0.1% TFA

Lysozym Protein

Retention time LacA bei 50% ACN

07 Integration 9721 - C:\MSDCHEM\200 Page 1  
 F10V8 10000.32m, C1200\THT-Peapen, ml\_3 16084 1998-08-27/20:13  
 Signal: 10000.32m, C1200\THT-Peapen, ml\_3 16084 1998-08-27  
 Method: 10000.32m, C1200\THT-Peapen, ml\_3 16084 1998-08-27  
 Sep. M/Sec: 25/1 Control: 10000.32m, C1200\THT-Peapen, ml\_3 16084 1998-08-27  
 Sample Type: Integration Signal: 10000.32m, C1200\THT-Peapen, ml\_3 16084 1998-08-27  
 Acquisition: 1998-08-27/20:13 Report: 10000.32m, C1200\THT-Peapen, ml\_3 16084 1998-08-27  
 Method: 10000.32m, C1200\THT-Peapen, ml\_3 16084 1998-08-27 Weight: 1.00000



Glykische Proteinierung:

Lys, LACB, 238

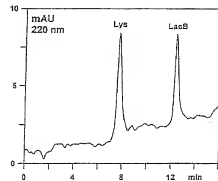
30-60% ACN / 15 min, 0.1% TFA

4.5 / 13000 Lac

215 nm

A790825-36

No.	Ret.	Time	Type	Area	Height	Height	Width	Class
1	1.43	1.43	1.43	1.43	1.43	1.43	1.43	44
2	8.45	8.45	8.45	8.45	8.45	8.45	8.45	1156
3	13.79	13.79	13.79	13.79	13.79	13.79	13.79	2080



Separation of proteins in a monolithic capillary column

Column, PS-DVB (monolith, 100 x 0.32 mm); chromatographic conditions, mobile phase, (A) H<sub>2</sub>O, 0.1% TFA, (B) ACN, 0.1% TFA, linear gradient, 30-60% B in 15 min; flow rate, 4.5 µl min<sup>-1</sup>; temperature, 25 °C; detection, UV, 220 nm; sample, lysozyme, β-lactoglobulin B, 20 ng each.

Kanal eines Trennung von Oligonucleotiden in Verdünnung 1:13.5

1:13.5

$l = 82 \text{ mm}$ ,  $id = 200 \mu\text{m}$

Fluss: A: 50 mM TEAA pH 6.8

B: 50 mM TEAA 20% ACN pH 6.8

Temperatur: 50°C

Spültemp. TSO 85375, 6 min

Flow 120 / 3.3, 11 min / 946

Welle: APP 0209.5711

Trennung von  $dt_8$ ,  $dt_{16}$

Peakhöhe mit Gradient 0-10% ACN/min. 0.11 min  
= 6.65

Trennung mit  $dt_{12-18}$

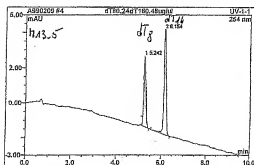
verschiedene Gradienten Kurven.

gute Trennung: 30-50% B/10 min

6-10% ACN/10 min

Operator: 72551 Thermo: A990209 Sequence: A990209 Page 4/1  
10.2.1999 2:58 PM

4 dt12-18 0.43 g/gly  
5-100% B/10 min A: 50 mM TEAA pH 6.8 B: 50 mM TEAA 20% ACN pH 6.8 TSO 85375, 6 min TSO 85375, 6 min  
Sample Name: dt12-18 0.43 g/gly Volume: 25.0  $\mu\text{L}$   
Chemical Program: Chemac UPL-1  
Queue Method: C15011 Recording Time: 44.07.58 19.00



No.	Ret. Time	Area	Height	Half Width	Plate	Asymmetry
	min	mm <sup>2</sup>	mAU	min	(RT)	(AU)
1	5.342	0.460	4.000	0.150	15000	1.200
2	5.812	0.744	0.000	0.155	18000	1.231
Total		1.194	10.001			

Operator: 72551 Thermo: A990209 Sequence: A990209 Page 11/1  
10.2.1999 2:54 PM

11 dt12-18 0.33 g/gly  
20-60% B/10 min A: 50 mM TEAA pH 6.8 B: 50 mM TEAA 20% ACN pH 6.8 TSO 85375, 6 min TSO 85375, 6 min  
Sample Name: dt12-18 0.33 g/gly Volume: 25.0  $\mu\text{L}$   
Chemical Program: Chemac UPL-1  
Queue Method: C15011 Recording Time: 09.01.09 2:12.58

